REVIEW



In vitro fertilization represents a risk factor for vasa praevia

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Abstract

Vasa praevia is a rare but very dangerous obstetrical condition. The purpose of our article is to evaluate data available in literature that indicate in vitro fertilization as a risk factor for vasa praevia. PubMed Library and Cochrane Database were searched using the keywords vasa praevia, in vitro fertilization, velamentous cord insertion, placenta praevia. The conditions related to in vitro fertilization that increase the risk of vasa praevia formation were identified and discussed. Also, the diagnosis and management options were reviewed. In vitro fertilization represents a risk factor for vasa praevia and all such pregnancies should be screened by transvaginal ultrasound for vasa praevia.

Keywords: vasa praevia, in vitro fertilization, velamentous cord insertion, placenta praevia.

→ Introduction

Vasa praevia is an obstetric condition that is associated with significant perinatal mortality and morbidity [1–6]. It occurs when the umbilical cord blood vessels are exposed within the placental membranes between internal os of the cervix and presenting part of the fetus [5, 7–9]. The importance of this pathology is underlined by its most feared complication, fetal death by exsanguinations, which occurs in more than 60% of the prenatally undetected cases [4, 5, 10, 11]. Though the approximate incidence of vasa praevia is considered to be one in 2500 pregnancies [12, 13], the true incidence is not known, with a very wide range from one in 10 000 to one in 5200 pregnancies [13, 14] reported in literature. The incidence is considered much higher, one in 365 [15] to one in 700, among patients who conceive through assisted reproductive technologies [16–18]. The most important risk factors for vasa praevia are umbilical cord and placental abnormallities, which are more common in pregnancies achieved after in vitro fertilization (IVF) [17, 19]. Since the number of IVF pregnancies has been increasing constantly over the last decade, a higher incidence of vasa praevia related complications may be encountered in the near future. The main objective of this article is to review data available in literature that indicate IVF as a risk factor for vasa praevia. We found no systematic review of the conditions related to IVF pregnancies that are considered risk factors for vasa praevia in English literature.

Our review consisted in a search of articles published in English. PubMed Library and Cochrane Database were searched for relevant articles including clinical trials, reviews, guidelines and case reports using the key words vasa praevia, in vitro fertilization, velamentous cord insertion. The websites of the International Vasa Praevia

Foundation and the UK Vasa Praevia Raising Awareness Organization were also reviewed for links to literature that may not have been indexed in medical databases. Articles assessing the ultrasound diagnosis of vasa praevia and the strategy of obstetrical management were included. A number of 204 articles were found. Most of case report articles discuss one up to three cases of vasa praevia and focus on the ultrasound criteria for diagnosis and the obstetrical management. Actually, they do not add much to the body of knowledge but they underline the severe potential of the situation. A review of the articles title and abstracts for relevance regarding our topic resulted in a number of 89 articles for review.

☐ Risk factors for vasa praevia

Two main types of *vasa praevia* are described: type I with velamentous insertion of the umbilical cord and type II with bilobed or succenturiate placenta [20–22]. In a velamentous insertion, the umbilical cord inserts directly into the membranes through which unprotected vessels then run until they end in the placenta. In type II *vasa praevia*, exposed vessels run through the membranes between lobes of a bilobed placenta [20, 22, 23].

The most important risk factors for *vasa praevia* are velamentous cord insertion [2, 19, 24], second-trimester low-lying placenta or *placenta praevia* [2, 10, 19, 25–27], pregnancies conceived after use of assisted reproductive technologies [2, 10, 19, 27], bilobed and succenturiate lobe placentas situated in the lower uterine segment [2, 10], and multiple pregnancies [2, 10, 14, 17, 19, 27–30]. Considering the risk factors, many authors consider that anomalies of placenta and umbilical cord insertion are prerequisite for *vasa praevia* [2, 8, 10, 19, 24–27, 31, 32].

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□ In vitro fertilization is related to higher incidence of placental abnormalities

Several authors link the abnormal placentation to assisted reproductive procedures, therefore IVF is considered a risk factor for vasa praevia [17, 32]. Early in 1984, a multi center study conducted by Jauniaux et al. investigated the pathologic features of placentas from singleton pregnancies obtained by IVF and embryo transfer (IVF-ET) [33]. They collected and examined 100 placentas from IVF pregnancies and found that the incidence of bilobate and succenturiate placenta was 22% in the IVF group compared to 6% in the control group. Results were considered statistically significant (p<0.5). Their conclusions were confirmed by the study performed by Englert et al. that evaluated the macroscopic characteristics of 100 fetal adnexa from pregnancies obtained by IVF-ET and compared with data for normal pregnancies taken from the literature. Material was obtained from 63 singleton, 15 twin, one triplet and one quadruplet pregnancies. They found normal placental morphology but abnormal insertion of the umbilical cord. Marginal (15%) and velamentous (14%) insertions of the umbilical cord were found more frequently than in a general obstetrical population (6% and 1%, respectively). Excluding placentae from multiple pregnancies (which are known to have a higher incidence of abnormal cord insertion), the frequency did not decrease and remained significantly higher than in a normal population (p<0.01 and p<00.001, for marginal and velamentous insertion respectively [34]. Schachter et al. (2002) studied a total of 72 818 deliveries, from 1987 to 2001; 1173 of them resulting from IVF pregnancies, and found 12 cases of vasa praevia. The overall incidence of vasa praevia was 1:6068 deliveries, similar to reports by other authors and 1:293 among the IVF pregnancies [17]. Romundstad et al. (2006) underlined the risk of placental abnormalities in pregnancies following assisted reproductive technology by comparing the incidence of placenta praevia following spontaneous and IVF pregnancies in the same mother. The study identified 1349 women who had conceived both naturally and after assisted fertilization in Norway, between 1988 and 2002. They found that the risk of placenta praevia was nearly three-fold higher in the pregnancy following assisted fertilization [adjusted odds ratio (OR) 2.9, 95% confidence interval (CI) 1.4–6.1], compared with that in the naturally conceived pregnancy. The study concluded that IVF is associated with an increased risk of placenta praevia that may be caused by factors related to the reproductive technology [18].

High levels of estradiol are related to abnormal placentation

The mechanism triggered by the IVF that leads to abnormal placentation is not completely understood [18, 34]. The high levels of estradiol present during the IVF stimulation cycle may interfere with normal placentation due to the stimulating effect on the endometrium. The transfer of a fresh embryo on the same cycle after oocyte retrieval means that implantation will take place in the context of high levels of estrogen due to prior ovarian stimulation. High estradiol concentrations at the time of

implantation may theoretically impair the endometrial response to trophoblast invasion, leading to abnormal placentation [35, 36]. Healy *et al.* (2010) have found that obstetric hemorrhage caused by *placenta praevia* and placental abruption are more frequent in IVF pregnancies and have suggested that a possible mechanism is the effect of high estradiol concentrations on the endometrium at the time of implantation [37]. Farhi *et al.* (2010) investigated a possible association between high estradiol concentrations and abnormal placentation by assessing the number and rate of pregnancy complications related to abnormal placentation. They found that the high estradiol concentration group of >10 000 pmol/L had significantly more complications related to abnormal placentation [35].

→ Abnormal cord insertion in IVF

Velamentous cord insertion represents the insertion of the umbilical cord into the membranes away from the placental margin. This results in the umbilical vessels lacking the protection of Wharton's jelly for the section between the insertion of the umbilical cord and the placental margin. Velamentous insertion of the umbilical cord is associated with vasa praevia [8, 38]. There are three possible theories that approach the etiopathogeny of velamentous cord insertion and vasa praevia: (1) an initial normal insertion of the umbilical cord can turn into a velamentous one due to the regress of the surrounding chorion frondosum caused by the placental expansion; (2) velamentous insertion of the cord favors the formation of big vessels extending to the margin of the placenta; (3) abnormal morphology of the placenta may be consecutive to distorted uterine anatomy such as myomas, uterine malformations and septa [30, 39–41].

Jauniaux et al. (1990) investigated placental shape and umbilical cord insertion among pregnancies obtained by IVF [33, 42]. The distance between cord insertion and placental margin was measured and umbilical cords inserted at less than 2 cm from the placental margin were considered marginal. The incidence of marginal and velamentous cord insertion was 26% and 12% in the IVF group compared to respectively 10% and 2% in the control group. These findings are confirmed by Schachter et al. who assessed a total of 72 818 deliveries and found a incidence of velamentous cord insertion of 1:743 in non-IVF pregnancies compared to 1:167 in the IVF group [17]. Ebbing et al. (2013) performed a population based study of 634 741 pregnancies between 1999–2009, aiming to determine the prevalence and the risk factors for anomalous insertions of the umbilical cord. They found that the prevalence of velamentous and marginal insertions of the umbilical cord was 7.8% in singletons and 16.9% in twin gestations, with marginal insertion being more common than velamentous [38, 43]. Delbaere et al. (2007) conducted the largest study of umbilical cord anomalies after IVF procedures. The study included over 4000 twin pregnancies, between 1995-2004, and concluded that umbilical cord anomalies are more frequent in twins after assisted reproduction and are influenced by the used technique. In twins conceived after IVF, the incidence of velamentous cord insertion was 7.4%, and after intracytoplasmatic sperm injection (ICSI), where a single sperm is injected into an egg, it was 10.4% [44].

A very rare case umbilical cord anomaly after IVF was reported by Canda *et al.* (2013). A patient with unicornuate uterus that achieved pregnancy on the fourth IVF attempt was diagnosed with velamentous and furcate cord insertion with placenta accreta [45]. Another very interesting case was reported by Hasegawa *et al.* (2011). They examined an IVF pregnancy at eight weeks of gestation and found the umbilical cord insertion with a viable fetus located on the septum membrane of dichorionic twin pregnancy, while the other fetus was observed to have vanished. Velamentous cord insertion with long membranous umbilical vessels was notice at delivery [46].

In conclusion, IVF pregnancies are related to higher incidence of multiple gestations [47–56] and abnormal insertion of the umbilical cord [17, 34, 38, 43, 44]. Both conditions are prerequisite for *vasa praevia* [12, 39, 40, 57].

Differences between frozen and fresh embryo transfer

Initially considered a strategy to reduce the rate of ovarian hyperstimulation syndrome, embryo freezing and transfer in another cycle seems to offer even more benefits than the fresh embryo transfer [58].

A preliminary study conducted by Imudia et al. (2013) suggests that elective cryopreservation of all embryos in patients with elevated peak serum of estradiol for subsequent cryopreservation and embryo transfer in cycles with a better physiologic hormonal milieu may reduce the odds of small for gestational age newborns and preeclampsia in IVF singleton deliveries [59]. Their results are confirmed by Korosec et al. (2014), who investigated the outcomes of singleton pregnancy after IVF with fresh or frozen embryo transfer and also the incidence of placenta praevia. They found that placenta praevia rates are lower in the frozen embryo transfer group, and the newborns had higher gestational weight than in the fresh embryo transfer group [60]. Unfortunately, they did not include the serum estradiol measurements into the analysis. A multicenter, prospective, randomized controlled clinical trial that aims to demonstrate that elective embryo cryopreservation and frozen-thawed embryo transfer will reduce the incidence of pregnancy complications related to placental abnormalities and increase the rate of live births in patients who need IVF to achieve pregnancy was initiated in 2014 by Shi et al. Their results are expected to make an impact in embryo transfer strategy [61].

☐ Multiple pregnancy vs. IVF vs. vasa praevia

Over 20% of all deliveries resulting from IVF/intracytoplasmic sperm injection (ICSI) include more than one fetus [43, 62]. One of the major obstacles in IVF remains the high twin birth rate and the complications related to multiple gestations. These problems can be solved by implementing elective single-embryo transfer (eSET), diminishing the twin birth rate without affecting the overall goal of achieving a healthy infant [47, 56, 63].

₽ Prenatal diagnosis in first or second trimester scan?

Vasa praevia can be diagnosed antenatally using combined abdominal and transvaginal ultrasound and color flow mapping; however, many cases are not diagnosed [64]. The cases not diagnosed antenatally are related to severe complication such as fetal death, low Apgar scores and severe anemia [2, 8, 64–66]. In 1801, the first case of ruptured vasa praevia was described in literature [67]. The first report of ultrasonographic diagnosis of vasa praevia appeared in the literature in 1987 [68]. Several authors report cases of vasa praevia diagnosed using color Doppler [3, 5, 6, 9, 10, 19, 65, 69, 70]. Transvaginal ultrasound is considered extremely important for an accurate diagnostic [2, 4, 16, 39]. Some authors emphasize the importance of tree-dimensional ultrasonography in establishing the diagnosis [20, 71–74]. There are two very important ultrasound exams during the follow-up of a pregnancy: the first trimester morphology scan and the second trimester morphology scan. These should be performed by experienced practitioners and could represent the ideal moment for the diagnosis of vasa praevia [28, 75]. Hasegawa et al. (2011) assessed the usefulness for predicting vasa praevia by detecting a cord insertion site in the lower third of the uterus between 9 and 13 weeks gestation and concluded that ultrasound screening in the first trimester of the cases with low cord insertion is effective for the detection of vasa praevia [76].

Sepulveda (2006) undertook a prospective study and screened 533 consecutive pregnancies during the nuchal translucency scan (11–14 weeks) for velamentous cord insertion. They followed all cases until delivery and found that no case of velamentous cord insertion was missed at the first trimester scan. Their study indicated that the diagnosing velamentous cord insertion at the nuchal scan in the first trimester is possible and recommendable [77].

Several authors reported cases of *vasa praevia* diagnosed in the second trimester [13, 28, 39, 75].

Canterino *et al.* (2005) reported a case of *vasa praevia* where 3D sonography with power Doppler angiography was used in order to certify the diagnosis [72].

Cipriano *et al.* analyzed the cost-effectiveness of targeted and universal screening for *vasa praevia* at 18–20 weeks of gestation in singleton and twin pregnancies. They found that screening with transvaginal ultrasound and color Doppler for IVF pregnancies or when the placenta has been found to be associated with one or more risk factors is cost-effective. The same screening in all population is not cost-effective [78].

Management of diagnosed cases

The outcome of a pregnancy with is *vasa praevia* is mainly determined by the early recognition of the pathology. Accurate prenatal diagnosis and Cesarean delivery before rupture of the membranes is associated with a 97% survival rate [11–13, 16, 40]. When the diagnosis is made antenatal, the safest form of delivery is elective Caesarean prior to the onset of labor [39, 79]. Consideration should be given to hospitalization at about 30 to 32 weeks and administration of corticosteroids to

promote fetal lung maturation [39, 80]. The optimal gestational age at delivery is difficult to establish [81]. The largest published series suggests that delivery by elective Caesarean section at 35 to 36 weeks gestation, prior to the formation of lower uterine segment is reasonable, thereby avoiding the risk of membrane rupture and fetal exsanguinations [7, 29, 39, 80, 82]. Several authors reported cases where delivery was delayed after 36 weeks but the risk of spontaneous rupture of membranes [83] and fetal exsanguination must be kept in mind [80].

Fetal therapy *in utero* may represent a solution in the near future [84–86]. The case of patient who underwent successful laser photocoagulation of a type II *vasa praevia* at 32 weeks gestation and subsequently delivered vaginally at term without complications is reported in literature [86].

☐ The prospect of this pathology in the near future

There is an increasing demand for assisted reproductive technology nowadays. Due to present social context regarding the age when women choose to procreate this trend is more likely to continue. Stress is a very common feature among women that undergo IVF procedures and this can also favor the development of placental abnormalities [87]. The increasing number of pregnancies obtained by IVF will make "niche" pathology such as vasa praevia more common for the current obstetrician. Although there are many reports in literature that indicate that IVF is a risk factor for vasa praevia, the exact mechanism of vasa praevia formation is not completely understood. The study of the placentas using immunohistochemistry could offer some answers about the placental vascular changes related to IVF pregnancies [88]. A very interesting idea was to compare the incidence of placental abnormalities in the same mother having both kinds of pregnancy (spontaneous and IVF). This eliminates most of the individual related factors and highlights the risk induced by artificial reproduction technology. Our review identifies the possible etiopathogenic paths described in literature and underlines that IVF generates a complex of factors that favor vasa praevia formation. High estrogen levels at the time of implantation and the transfer of more than one embryo may induce anomalies of placentation and umbilical cord implantation and therefore the formation of vasa praevia. Our review also suggests that the transfer of a single frozen embryo in another cycle may reduce the rate of abnormal placentation and umbilical cord implantation. The transfer of a single embryo versus two embryos remains a disputed subject between IVF centers nowadays, with the balance shifting towards single embryo transfer in most centers [51, 56, 89]. Since the morbidity of this condition is mostly determined by the lack of recognition, a detailed ultrasound screening should be performed in all IVF pregnancies [32]. The transvaginal scan can identify the presence of vasa praevia or its high risk factors, such as velamentous cord insertion and abnormal placenta, as early as the first trimester nuchal scan. The diagnosed cases should be monitored closely and cesarean section should be scheduled prior to labor onset.

Awareness of the risk factors, diagnosis and management of *vasa praevia* needs to be raised among obstetricians. Our article highlights that IVF represents a risk factor for *vasa praevia*, and all IVF pregnancies should be screened by transvaginal ultrasound for *vasa praevia*.

Conflict of interests

The authors declare that they have no conflict of interests.

References

- [1] Javid N, Sullivan EA, Halliday LE, Duncombe G, Homer CS. "Wrapping myself in cotton wool": Australian women's experience of being diagnosed with vasa praevia. BMC Pregnancy Childbirth, 2014, 14:318.
- [2] Carnide C, Jerónimo M, Faria D, Silva IS. Twin pregnancy complicated by vasa previa. BMJ Case Rep, 2012 Dec 14, 2012:bcr2012006484.
- [3] Rao KP, Belogolovkin V, Yankowitz J, Spinnato JA 2nd. Abnormal placentation: evidence-based diagnosis and management of placenta previa, placenta accreta, and vasa previa. Obstet Gynecol Surv, 2012, 67(8):503–519.
- [4] Smorgick N, Tovbin Y, Ushakov F, Vaknin Z, Barzilay B, Herman A, Maymon R. Is neonatal risk from vasa previa preventable? The 20-year experience from a single medical center. J Clin Ultrasound, 2010, 38(3):118–122.
- [5] Pérez Rodríguez MJ, de Frutos Moneo E, Nieto Llanos S, Clemente Pollán J. Vasa praevia rupture in velamentous insertion of the umbilical cord: the importance of prenatal diagnosis. An Pediatr (Barc), 2014, 81(6):393–395.
- [6] Clerici G, Burnelli L, Lauro V, Pilu GL, Di Renzo GC. Prenatal diagnosis of vasa previa presenting as amniotic band. 'A not so innocent amniotic band'. Ultrasound Obstet Gynecol, 1996, 7(1):61–63.
- [7] Bręborowicz GH, Markwitz W, Szpera-Goździewicz A, Dera-Szymanowska A, Ropacka-Lesiak M, Szymański P, Kubiaczyk-Paluch B. Prenatal diagnosis of vasa previa. J Matern Fetal Neonatal Med, 2015, 28(15):1806–1808.
- [8] Wiedaseck S, Monchek R. Placental and cord insertion pathologies: screening, diagnosis, and management. J Midwifery Womens Health, 2014, 59(3):328–335.
- [9] Araujo Júnior E, Filho HA, Pires CR, Zanforlin Filho SM, Moron AF. Prenatal diagnosis of vasa previa through color Doppler and three-dimensional power Doppler ultrasonography. A case report. Clin Exp Obstet Gynecol, 2006, 33(2):122–124.
- [10] Oyelese KO, Schwärzler P, Coates S, Sanusi FA, Hamid R, Campbell S. A strategy for reducing the mortality rate from vasa previa using transvaginal sonography with color Doppler. Ultrasound Obstet Gynecol, 1998, 12(6):434–438.
- [11] Oyelese Y, Catanzarite V, Prefumo F, Lashley S, Schachter M, Tovbin Y, Goldstein V, Smulian JC. Vasa previa: the impact of prenatal diagnosis on outcomes. Obstet Gynecol, 2004, 103(5 Pt 1):937–942.
- [12] Oyelese KO, Turner M, Lees C, Campbell S. Vasa previa: an avoidable obstetric tragedy. Obstet Gynecol Surv, 1999, 54(2):138–145.
- [13] Rebarber A, Dolin C, Fox NS, Klauser CK, Saltzman DH, Roman AS. Natural history of vasa previa across gestation using a screening protocol. J Ultrasound Med, 2014, 33(1): 141–147.
- [14] Lee W, Lee VL, Kirk JS, Sloan CT, Smith RS, Comstock CH. Vasa previa: prenatal diagnosis, natural evolution, and clinical outcome. Obstet Gynecol, 2000, 95(4):572–576.
- [15] Hasegawa J, Nakamura M, Ichizuka K, Matsuoka R, Sekizawa A, Okai T. Vasa previa is not infrequent. J Matern Fetal Neonatal Med, 2012, 25(12):2795–2796.
- [16] Kajimoto E, Matsuzaki S, Matsuzaki S, Tanaka Y, Kinugasa-Taniguchi Y, Mimura K, Kanagawa T, Kimura T. Challenges in diagnosis of pseudo vasa previa. Case Rep Obstet Gynecol, 2014, 2014:903920.
- [17] Schachter M, Tovbin Y, Arieli S, Friedler S, Ron-El R, Sherman D. *In vitro* fertilization is a risk factor for vasa previa. Fertil Steril, 2002, 78(3):642–643.

- [18] Romundstad LB, Romundstad PR, Sunde A, von Düring V, Skjaerven R, Vatten LJ. Increased risk of placenta previa in pregnancies following IVF/ICSI; a comparison of ART and non-ART pregnancies in the same mother. Hum Reprod, 2006, 21(9):2353–2358.
- [19] Aissi G, Sananes N, Veujoz M, Felder A, Kasbaoui SM, Trieu NT, Favre R, Nisand I. Vasa previa: of the diagnosis to neonatal prognosis. J Gynecol Obstet Biol Reprod (Paris), 2013, 42(6): 591–595.
- [20] Oyelese Y, Chavez MR, Yeo L, Giannina G, Kontopoulos EV, Smulian JC, Scorza WE. Three-dimensional sonographic diagnosis of vasa previa. Ultrasound Obstet Gynecol, 2004, 24(2):211–215.
- [21] Donnolley N, Halliday LE, Oyelese Y. Vasa praevia: a descriptive review of existing literature and the evolving role of ultrasound in prenatal screening. Australas J Ultrasound Med (AJUM), 2013, 16(2):71–76.
- [22] Catanzarite V, Maida C, Thomas W, Mendoza A, Stanco L, Piacquadio KM. Prenatal sonographic diagnosis of vasa previa: ultrasound findings and obstetric outcome in ten cases. Ultrasound Obstet Gynecol, 2001, 18(2):109–115.
- [23] Daly-Jones E, John A, Leahy A, Mckenna C, Sepulveda W. Vasa praevia; a preventable tragedy. Ultrasound, 2008, 16(1): 8–14.
- [24] Hasegawa J, Farina A, Nakamura M, Matsuoka R, Ichizuka K, Sekizawa A, Okai T. Analysis of the ultrasonographic findings predictive of vasa previa. Prenat Diagn, 2010, 30(12–13): 1121–1125.
- [25] Fung TY, Lau TK. Poor perinatal outcome associated with vasa previa: is it preventable? A report of three cases and review of the literature. Ultrasound Obstet Gynecol, 1998, 12(6):430–433.
- [26] Francois K, Mayer S, Harris C, Perlow JH. Association of vasa previa at delivery with a history of second-trimester placenta previa. J Reprod Med, 2003, 48(10):771–774.
- [27] Oyelese Y, Spong C, Fernandez MA, McLaren RA. Second trimester low-lying placenta and *in-vitro* fertilization? Exclude vasa previa. J Matern Fetal Med, 2000, 9(6):370–372.
- [28] Baulies S, Maiz N, Muñoz A, Torrents M, Echevarría M, Serra B. Prenatal ultrasound diagnosis of vasa praevia and analysis of risk factors. Prenat Diagn, 2007, 27(7):595–599.
- [29] Gandhi M, Cleary-Goldman J, Ferrara L, Ciorica D, Saltzman D, Rebarber A. The association between vasa previa, multiple gestations, and assisted reproductive technology. Am J Perinatol, 2008, 25(9):587–589.
- [30] Komatsu A, Kozuma S, Yoshida S, Hyodo H, Yamashita T, Kamei Y, Fujii T, Taketani Y. A case f vasa previa diagnosed prenatally, and review of the literature. J Med Ultrason (2001), 2011, 38(1):41–45.
- [31] Régis C, Mubiayi ND, Devisme L, Subtil D. Benckiser's hemorrhage: severe and inevitable? J Gynecol Obstet Biol Reprod (Paris), 2006, 35(5 Pt 1):517–521.
- [32] Al-Khaduri M, Kadoch IJ, Couturier B, Dubé J, Lapensée L, Bissonnette F. Vasa praevia after IVF: should there be guidelines? Report of two cases and literature review. Reprod Biomed Online, 2007, 14(3):372–374.
- [33] Jauniaux E, Englert Y, Vanesse M, Hiden M, Wilkin P. Pathologic features of placentas from singleton pregnancies obtained by in vitro fertilization and embryo transfer. Obstet Gynecol, 1990, 76(1):61–64.
- [34] Englert Y, Imbert MC, Van Rosendael E, Belaisch J, Segal L, Feichtinger W, Wilkin P, Frydman R, Leroy F. Morphological anomalies in the placentae of IVF pregnancies: preliminary report of a multicentric study. Hum Reprod, 1987, 2(2):155–157.
- [35] Farhi J, Ben-Haroush A, Andrawus N, Pinkas H, Sapir O, Fisch B, Ashkenazi J. High serum oestradiol concentrations in IVF cycles increase the risk of pregnancy complications related to abnormal placentation. Reprod Biomed Online, 2010, 21(3):331–337.
- [36] Simón C, Cano F, Valbuena D, Remohí J, Pellicer A. Clinical evidence for a detrimental effect on uterine receptivity of high serum oestradiol concentrations in high and normal responder patients. Hum Reprod, 1995, 10(9):2432–2437.
- [37] Healy DL, Breheny S, Halliday J, Jaques A, Rushford D, Garrett C, Talbot JM, Baker HW. Prevalence and risk factors for obstetric haemorrhage in 6730 singleton births after assisted reproductive technology in Victoria Australia. Hum Reprod, 2010, 25(1):265–274.

- [38] Ebbing C, Kiserud T, Johnsen SL, Albrechtsen S, Rasmussen S. Prevalence, risk factors and outcomes of velamentous and marginal cord insertions: a population-based study of 634,741 pregnancies. PLoS One, 2013, 8(7):e70380.
- [39] Gagnon R, Morin L, Bly S, Butt K, Cargill YM, Denis N, Hietala-Coyle MA, Lim KI, Ouellet A, Raciot MH, Salem S; Diagnostic Imaging Committee, Hudon L, Basso M, Bos H, Delisle MF, Farine D, Grabowska K, Menticoglou S, Mundle W, Murphy-Kaulbeck L, Pressey T, Roggensack A; Maternal Fetal Medicine Committee. Guidelines for the management of vasa previa. J Obstet Gynaecol Can, 2009, 31(8):748–760.
- [40] Oyelese Y, Smulian JC. Placenta previa, placenta accreta, and vasa previa. Obstet Gynecol, 2006, 107(4):927–941.
- [41] Stafford IP, Neumann DE, Jarrell H. Abnormal placental structure and vasa previa: confirmation of the relationship. J Ultrasound Med, 2004, 23(11):1521–1522.
- [42] Gavriil P, Jauniaux E, Leroy F. Pathologic examination of placentas from singleton and twin pregnancies obtained after in vitro fertilization and embryo transfer. Pediatr Pathol, 1993, 13(4):453–462.
- [43] de Mouzon J, Goossens V, Bhattacharya S, Castilla JA, Ferraretti AP, Korsak V, Kupka M, Nygren KG, Nyboe Andersen A; European IVF-monitoring (EIM) Consortium, for the European Society of Human Reproduction and Embryology (ESHRE). Assisted reproductive technology in Europe, 2006: results generated from European registers by ESHRE. Hum Reprod, 2010, 25(8):1851–1862.
- [44] Delbaere I, Goetgeluk S, Derom C, De Bacquer D, De Sutter P, Temmerman M. Umbilical cord anomalies are more frequent in twins after assisted reproduction. Hum Reprod, 2007, 22(10):2763–2767.
- [45] Canda MT, Demir N, Doganay L. Velamentous and furcate cord insertion with placenta accreta in an IVF pregnancy with unicornuate uterus. Case Rep Obstet Gynecol, 2013, 2013: 539379
- [46] Hasegawa J, Iwasaki S, Matsuoka R, Ichizuka K, Sekizawa A, Okai T. Velamentous cord insertion caused by oblique implantation after *in vitro* fertilization and embryo transfer. J Obstet Gynaecol Res, 2011, 37(11):1698–1701.
- [47] Pinborg A. IVF/ICSI twin pregnancies: risks and prevention. Hum Reprod Update, 2005, 11(6):575–593.
- [48] Allen VM, Wilson RD, Cheung A; Genetics Committee of the Society of Obstetricians and Gynaecologists of Canada (SOGC); Reproductive Endocrinology Infertility Committee of the Society of Obstetricians and Gynaecologists of Canada (SOGC). Pregnancy outcomes after assisted reproductive technology. J Obstet Gynaecol Can, 2006, 28(3):220–250.
- [49] Adashi EY, Barri PN, Berkowitz R, Braude P, Bryan E, Carr J, Cohen J, Collins J, Devroey P, Frydman R, Gardner D, Germond M, Gerris J, Gianaroli L, Hamberger L, Howles C, Jones H Jr, Lunenfeld B, Pope A, Reynolds M, Rosenwaks Z, Shieve LA, Serour GI, Shenfield F, Templeton A, van Steirteghem A, Veeck L, Wennerholm UB. Infertility therapy-associated multiple pregnancies (births): an ongoing epidemic. Reprod Biomed Online, 2003, 7(5):515–542.
- [50] Ombelet W, De Sutter P, Van der Elst J, Martens G. Multiple gestation and infertility treatment: registration, reflection and reaction – the Belgian project. Hum Reprod Update, 2005, 11(1):3–14
- [51] Sunderam S, Kissin DM, Crawford SB, Folger SG, Jamieson DJ, Barfield WD; Centers for Disease Control and Prevention (CDC). Assisted reproductive technology surveillance – United States, 2011. MMWR Surveill Summ, 2014, 63(Suppl 10): 1, 28
- [52] Kulkarni AD, Jamieson DJ, Jones HW Jr, Kissin DM, Gallo MF, Macaluso M, Adashi EY. Fertility treatments and multiple births in the United States. N Engl J Med, 2013, 369(23):2218–2225.
- [53] ***. Multiple gestation pregnancy. The ESHRE Capri Workshop Group. Hum Reprod, 2000, 15(8):1856–1864.
- [54] Pandian Z, Bhattacharya S, Ozturk O, Serour GI, Templeton A. Number of embryos for transfer following in-vitro fertilisation or intra-cytoplasmic sperm injection. Cochrane Database Syst Rev, 2004, (4):CD003416.
- [55] Sullivan EA, Zegers-Hochschild F, Mansour R, Ishihara O, de Mouzon J, Nygren KG, Adamson GD. International Committee for Monitoring Assisted Reproductive Technologies (ICMART) world report: assisted reproductive technology 2004. Hum Reprod, 2013, 28(5):1375–1390.

- [56] Scholten I, Chambers GM, van Loendersloot L, van der Veen F, Repping S, Gianotten J, Hompes PG, Ledger W, Mol BW. Impact of assisted reproductive technology on the incidence of multiple-gestation infants: a population perspective. Fertil Steril, 2015, 103(1):179–183.
- [57] Torrey WE. Vasa praevia. Am J Obstet Gynecol, 1952, 63: 146–152.
- [58] D'Angelo A, Amso N. Embryo freezing for preventing ovarian hyperstimulation syndrome. Cochrane Database Syst Rev, 2007, (3):CD002806.
- [59] Imudia AN, Awonuga AO, Kaimal AJ, Wright DL, Styer AK, Toth TL. Elective cryopreservation of all embryos with subsequent cryothaw embryo transfer in patients at risk for ovarian hyperstimulation syndrome reduces the risk of adverse obstetric outcomes: a preliminary study. Fertil Steril, 2013, 99(1):168–173.
- [60] Korosec S, Ban Frangez H, Verdenik I, Kladnik U, Kotar V, Virant-Klun I, Vrtacnik Bokal E. Singleton pregnancy outcomes after in vitro fertilization with fresh or frozen-thawed embryo transfer and incidence of placenta praevia. Biomed Res Int, 2014, 2014;431797.
- [61] Shi Y, Wei D, Liang X, Sun Y, Liu J, Cao Y, Zhang B, Legro RS, Zhang H, Chen ZJ1. Live birth after fresh embryo transfer vs elective embryo cryopreservation/frozen embryo transfer in women with polycystic ovary syndrome undergoing IVF (FreFro-PCOS): study protocol for a multicenter, prospective, randomized controlled clinical trial. Trials, 2014, 15:154.
- [62] Zegers-Hochschild F, Schwarze JE, Crosby JA, Musri C, do Carmo Borges de Souza M. Assisted reproductive technologies in Latin America: the Latin American Registry, 2012. Reprod Biomed Online, 2015, 30(1):43–51.
- [63] Ravhon A, Hurwitz A. Transfer of single embryo as a method to reduce twins pregnancy rate in *in-vitro* fertilization treatment. Harefuah, 2002, 141(3):301–305, 312.
- [64] Lijoi AF, Brady J. Vasa previa diagnosis and management. J Am Board Fam Pract, 2003, 16(6):543–548.
- [65] Meyer WJ, Blumenthal L, Cadkin A, Gauthier DW, Rotmensch S. Vasa previa: prenatal diagnosis with transvaginal color Doppler flow imaging. Am J Obstet Gynecol, 1993, 169(6):1627–1629.
- [66] Robert JA, Sepulveda W. Fetal exsanguination from ruptured vasa previa: still a catastrophic event in modern obstetrics. J Obstet Gynaecol, 2003, 23(5):574.
- [67] Lobstein J. Archives de l'art des accouchements. Strasbourg, 1801, 320 pp.
- [68] Gianopoulos J, Carver T, Tomich PG, Karlman R, Gadwood K. Diagnosis of vasa previa with ultrasonography. Obstet Gynecol, 1987, 69(3 Pt 2):488–491.
- [69] Nomiyama M, Toyota Y, Kawano H. Antenatal diagnosis of velamentous umbilical cord insertion and vasa previa with color Doppler imaging. Ultrasound Obstet Gynecol, 1998, 12(6):426–429.
- [70] Megier P, Desroches A, Esperandieu O, Mekari B. Prenatal diagnosis of vasa previa with velamentous cord insertion, using Doppler color echography. J Gynecol Obstet Biol Reprod (Paris), 1995, 24(4):415–417.
- [71] Lee W, Kirk JS, Comstock CH, Romero R. Vasa previa: prenatal detection by three-dimensional ultrasonography. Ultrasound Obstet Gynecol, 2000, 16(4):384–387.
- [72] Canterino JC, Mondestin-Sorrentino M, Muench MV, Feld S, Baum JD, Fernandez CO. Vasa previa: prenatal diagnosis

- and evaluation with 3-dimensional sonography and power angiography. J Ultrasound Med, 2005, 24(5):721–724; quiz 725.
- [73] Mabuchi Y, Yamoto M, Minami S, Boshi E, Yagi S, Oba N, Tanaka K, Umesaki N. Two cases of vasa previa diagnosed prenatally using three-dimensional ultrasonography. J Clin Ultrasound, 2010, 38(7):389–392.
- [74] Matijevic R, Kurjak A. The assessment of placental blood vessels by three-dimensional power Doppler ultrasound. J Perinat Med, 2002, 30(1):26–32.
- [75] Kanda E, Matsuda Y, Kamitomo M, Maeda T, Mihara K, Hatae M. Prenatal diagnosis and management of vasa previa: a 6-year review. J Obstet Gynaecol Res, 2011, 37(10):1391– 1396.
- [76] Hasegawa J, Nakamura M, Sekizawa A, Matsuoka R, Ichizuka K, Okai T. Prediction of risk for vasa previa at 9–13 weeks' gestation. J Obstet Gynaecol Res, 2011, 37(10):1346–1351.
- [77] Sepulveda W. Velamentous insertion of the umbilical cord: a first-trimester sonographic screening study. J Ultrasound Med, 2006, 25(8):963–968; quiz 970.
- [78] Cipriano LE, Barth WH Jr, Zaric GS. The cost-effectiveness of targeted or universal screening for vasa praevia at 18–20 weeks of gestation in Ontario. BJOG, 2010, 117(9):1108– 1118.
- [79] Hasegawa J, Arakaki T, Ichizuka K, Sekizawa A. Management of vasa previa during pregnancy. J Perinat Med, 2015, 43(6): 783–784.
- [80] Golic M, Hinkson L, Bamberg C, Rodekamp E, Brauer M, Sarioglu N, Henrich W. Vasa praevia: risk-adapted modification of the conventional management – a retrospective study. Ultraschall Med, 2013, 34(4):368–376.
- [81] Hoover MA, Allen A, La Rochelle F, Baig-Lewis S, Pilliod R, Caughey AB. Timing delivery of vasa previa: a decision analysis. Obstet Gynecol, 2014, 123(Suppl 1):148S–149S.
- [82] Robinson BK, Grobman WA. Effectiveness of timing strategies for delivery of individuals with vasa previa. Obstet Gynecol, 2011, 117(3):542–549.
- [83] Pirtea L, Grigoras D, Sas I, Pirtea M. Vasa previa. Gineco Ro, 2011, 7(3):146–148.
- [84] Quintero RA, Kontopoulos EV, Bornick PW, Allen MH. In utero laser treatment of type II vasa previa. J Matern Fetal Neonatal Med, 2007, 20(12):847–851.
- [85] Chmait RH, Chavira E, Kontopoulos EV, Quintero RA. Third trimester fetoscopic laser ablation of type II vasa previa. J Matern Fetal Neonatal Med, 2010, 23(5):459–462.
- [86] Johnston R, Shrivastava VK, Chmait RH. Term vaginal delivery following fetoscopic laser photocoagulation of type II vasa previa. Fetal Diagn Ther, 2014, 35(1):62–64.
- [87] Marinescu IP, Foarfă MC, Pîrlog MC, Turculeanu A. Prenatal depression and stress – risk factors for placental pathology and spontaneous abortion. Rom J Morphol Embryol, 2014, 55(3 Suppl):1155–1160.
- [88] Gheorman V, Gheorman L, Ivănuş C, Pană RC, Gogănău AM, Pătraşcu A. Comparative study of placenta acute fetal distress and diabetes associated with pregnancy. Rom J Morphol Embryol, 2013, 54(3):505–511.
- [89] Fechner AJ, Brown KR, Onwubalili N, Jindal SK, Weiss G, Goldsmith LT, McGovern PG. Effect of single embryo transfer on the risk of preterm birth associated with in vitro fertilization. J Assist Reprod Genet, 2015, 32(2):221–224.

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